

# **MICRO**

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**RR-8.0**

# Contents

General Microbiology	1
Hospital Infection Control	7
Immunology	10
Laboratory Diagnosis of Infections : Bacterial and Fungal	15
Laboratory Diagnosis of Infections : Viral and Parasitic	18
Central Nervous System	23
Cardiovascular System : Infections of Heart	27
Cardiovascular System : Bloodstream and Lymphatic Infections	30
Respiratory System	38
Gastrointestinal and Hepatobiliary System	45
Genitourinary System	51
Skin, Subcutaneous and Musculoskeletal System	53
Miscellaneous Infections	60



# GENERAL MICROBIOLOGY

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Kary B mullis :

Invented PCR (molecular) : Detects gene.

Eva Engvall (Along with Peter Perlman) :

Developed ELISA (Immunological) : Detects Ag/Ab.



## Microscopy & Staining

00:09:58

### MICROSCOPY

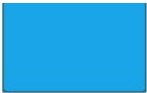
Detection of organism → morphology : Staining (killed).  
→ motility : Hanging drop (Live).

Types of microscopes to visualize motility :

	Bright field microscope (Light)	Darkfield microscope	Phase contrast microscope
Features	m/c used	Darkfield (Special condenser) > Phase contrast (Phase plate)	
Characteristics	Bright organism with bright background	 Bright organism with dark background	 Dark organism with bright background
motility Observed	All <b>except thin</b> organisms	Thin organisms motility visualized (Spirochetes : <b>Corkscrew</b> motility)	

Darkfield microscope : Contains **dark field condensor**.

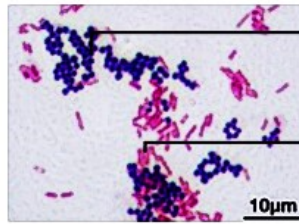
Phase contrast microscope : Contains **phase plate**.



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**STAINING**

**Gram Stain :**



Violet : Gram Positive

Pink : Gram Negative

Smear preparation :

Primary staining : Crystal violet/Gentian/methyl violet

(1 min)

↓ wash



Organisms stain violet

Add mordant : Gram's Iodine

(1 min)

↓ wash



Organisms remain violet

Add decolorization (Critical step) : Acetone (2-3 secs)/

(Few seconds)

Alcohol (20-30 secs)

↓ wash



Decolorize ⊖ : Retain 1<sup>o</sup> stain.  
Decolorize ⊕ : Colorless.

Counter staining : Saffranin/Dilute carbol fuschin

(1 min)



Violet : Gram ⊕ (Thick cell wall)  
Pink : Gram ⊖ (Thin cell wall)

**Acid Fast Stain/Ziehl-Neelsen Stain :**



Blue background

Pink : Acid fast

No pink : Non-acid fast

Smear Preparation :

Primary staining + mordant : Concentrated Carbol fuchsin (Pink)

(5 min)

+ Intermittent heating



↓ wash

Decolorization (Critical step) : H<sub>2</sub>SO<sub>4</sub>

(1 min)

↓ wash

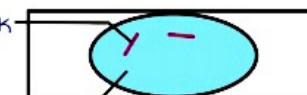
Decolorize ⊖ ← Pink  
Decolorize ⊕ ← Colourless



Counter staining : methylene blue

Acid fast organism ← Pink  
(Cell wall : mycolic acid)

Non-acid fast organisms ← Blue  
(Not visible against same blue background)

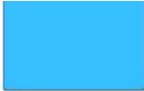


Note :

Concentration of H<sub>2</sub>SO<sub>4</sub>

- 20% : m. Tb
- 5% : m. leprae.
- 1% : Nocardia & parasites.





Culture & Molecular Methods

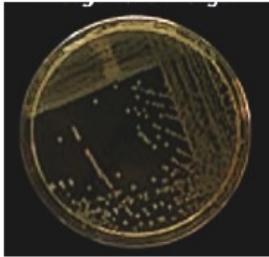
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Culture media (Cm) :

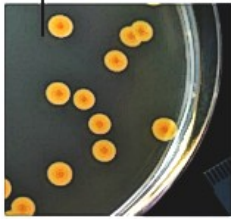
Simple/Basal Cm :

- Nutrient agar (Straw-colored).
- Appearance : white-grey colonies.
- Exceptions :



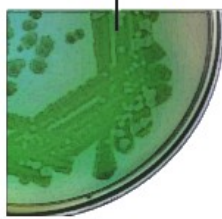
white-grey colonies

Golden yellow pigment



Staphylococcus aureus

Bluish-green pigment



Pseudomonas aeruginosa

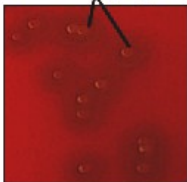
Enriched media :

- Blood agar.
- Normal Appearance : No zone surrounding colonies.
- Exceptions :



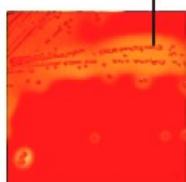
No zone

α-hemolysis (Partial) :  
Greenish-grey zone



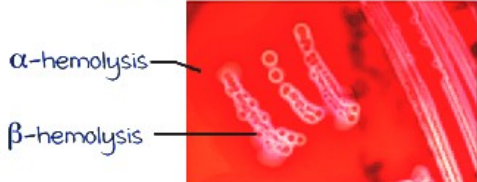
Pneumococcus

β-hemolysis (Complete) :  
Clear zone



Staphylococcus

Double zone/Target hemolysis :  
Complete followed by partial hemolysis



α-hemolysis

β-hemolysis

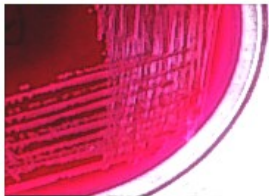
Clostridium perfringens

Differential media :

- McConkey agar :

Only gram -ve organisms grow.

Pink colonies : Lactose fermenters (LF)



← E. coli, Klebsiella →

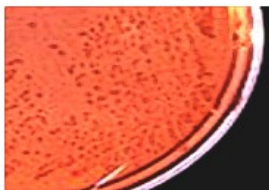
- CLED (Cystine-lactose-electrolyte deficient) agar :

Gram (-), Gram (+) & Candida grows.

Yellow colonies (LF)



Pale colonies : Non-lactose fermenters (NLF)



← many organisms →

Blue colonies (NLF)



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Enrichment CM :

- use : Inhibit commensals in stool sample.
- Types :

	Selenite F broth	Tetrathionate broth	Alkaline peptone water
Colour of CM	Light brown	Bluish green	Straw colour
Pathogen	Salmonella, Shigella		Vibrio

Note :

NALC-NaOH method : Inhibits commensals in sputum sample

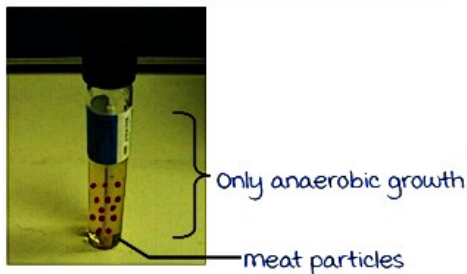
- N-Acetyl-L-cysteine : Liquefies sputum.
- NaOH : Inhibit commensals.

Transport media :

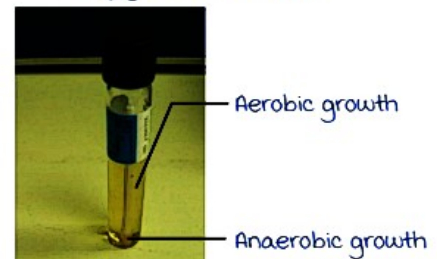


Anaerobic CM :

- Robertson cooked meat (RCM) : Best


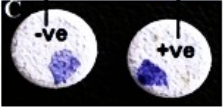


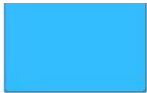
- Thioglycollate broth :



Culture Identification :

Biochemical Tests :

	Catalase Test	Oxidase Test	Urease Test
Features	Bubble No bubble 	No blue colour Blue colour 	<ul style="list-style-type: none"> <li>• Pink color : urease +ve</li> <li>• No pink color : urease -ve</li> </ul>
Organisms	Catalase -ve : <ul style="list-style-type: none"> <li>• Streptococcus</li> <li>• Pneumococcus</li> <li>• Enterococcus</li> </ul>	Oxidase +ve : <ul style="list-style-type: none"> <li>• Vibrio</li> <li>• Pseudomonas</li> <li>• Campylobacter</li> <li>• Helicobacter</li> </ul>	Urease +ve : <ul style="list-style-type: none"> <li>• Helicobacter</li> <li>• Proteus</li> </ul>



**MOLECULAR METHOD**

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**PCR :**

Steps :

Nucleic acid extraction : Enzyme method (Add Lysozyme).

Nucleic acid amplification in **thermocycler** →

- Denaturation (95°C) : ds to 2 single strands.
- Annealing (55°C) : Primer attachment.
- Extension (72°C) : Primer extension.

Nucleic acid detection : Gel electrophoresis/Fluorescent method.

Uses :

- Diagnostic test : Detects gene.
- **Prognostic test** : To monitor Rx response.
  - Gives organism load.
  - If PCR ⊕ : If ↑CT (Cycle Threshold) value → ↓Organism → ↑Prognosis.

modifications :

- Conventional PCR : Detects only DNA.
- Real-time RT (Reverse Transcriptase) PCR :
  - Detects DNA & RNA.
  - Semiautomated.
- Automated Realtime RT-PCR : **CBNAAT** (Cartridge based) & **TRUNAT** (Chip based).
- Multiplex Realtime RT-PCR : Detects multiple organisms.

**Antimicrobial Susceptibility Testing (AST)**

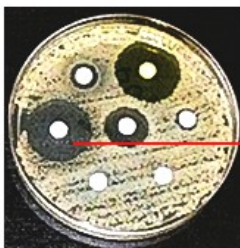
01:00:40

**Phenotypic method :**

Culture method.

Kirby Bauer disk diffusion : **m/c.**

- Mueller Hinton Agar (MHA).
- Zone of inhibition → Present : Antibiotic sensitive.  
→ Absent : Antibiotic resistant.
- Minimum Inhibitory Concentration (MIC) of antibiotic not obtained.



Zone of inhibition

Antibiotic disk

E-strip :

- MHA.
- **MIC obtained.**



Zone of inhibition : Antibiotic sensitive

MIC

Antibiotic strip

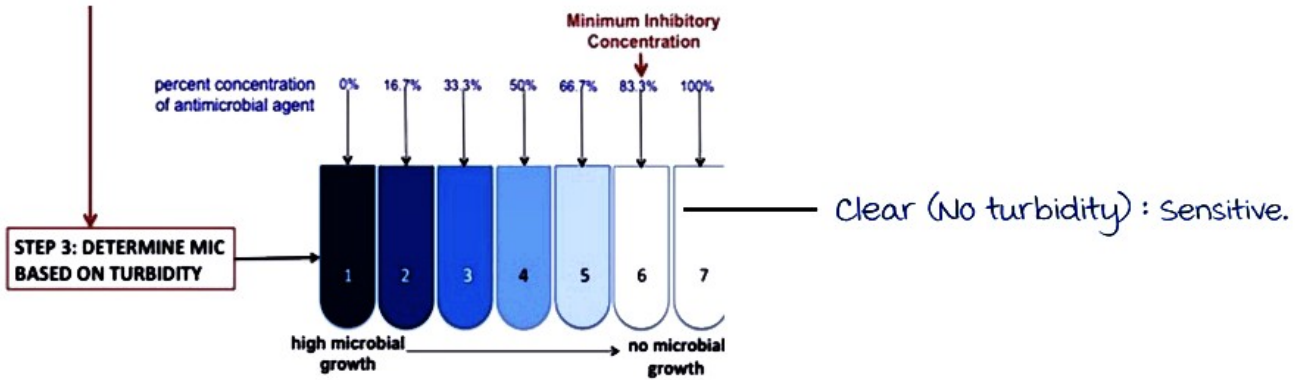




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Broth dilution : Gold standard

- mueller Hinton Broth (MHB).
- MIC obtained.



Genotypic method :

molecular method.

PCR :

Detects resistant gene → If present : Antibiotic resistant.  
 → If absent : Antibiotic sensitive.

Example : CBNAAT detects → m. Tb  
 → rpo-β gene (If ⊕ : Rifampicin resistance).



# HOSPITAL INFECTION CONTROL

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## Hand Hygiene & PPE

00:00:42

### Hand Hygiene :

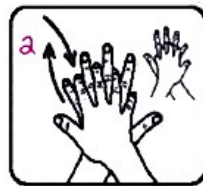
methods :

1. Hand wash	2. Hand rub
<ul style="list-style-type: none"> <li>• Better method.</li> <li>• Soap &amp; water used.</li> </ul>	<ul style="list-style-type: none"> <li>• If visibly soiled : Not acceptable.</li> <li>• Disinfectant used.</li> </ul>

Steps :



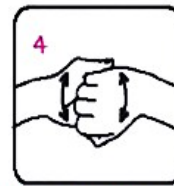
1 Rub hands palm to palm.



2 Rub hand's back with other palm, fingers interlaced.



3 Rub palm to palm, fingers interlaced.



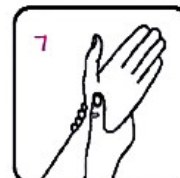
4 rub backs of fingers to opposing palms, hands clasped.



5 Rub tips of fingers onto palm in a circular motion.



6 Rub each thumb with rotational motion.



7 Rub each wrist.

5 moments for hand hygiene 🚩 :

- Before & after touching patient.
- Before & after procedures & fluid exposure.
- After touching patient surroundings.

### Personal Protective Equipment (PPE) :

Essential PPE : Gown, gloves, mask, goggles & faceshield.

Two methods :

	1. Donning	2. Doffing
Action	🚩 Putting PPE on	Removing PPE
Sequence	Gown → mask → Goggles/Face shield → Gloves.	Gloves → Goggles/Face shield ⇔ Gown → mask

----- Active space -----

**Biomedical Waste Management (BMW)** 🚮

00:11:40

method of **segregation** & **disposal** of waste contaminated with **microorganisms**.

	Yellow Bag	Red Bag	White Bag	Blue Bag
Type of Waste	<ul style="list-style-type: none"> <li>Cotton, Linen</li> <li>Tissues, culture media</li> <li>Chemicals, medicines</li> <li><b>Blood bags (Plastic)</b></li> </ul>	<ul style="list-style-type: none"> <li>Plastic : <b>Urine bag</b></li> <li>Rubber</li> </ul>	metallic Sharps	<ul style="list-style-type: none"> <li>metals</li> <li>Glass</li> </ul>
Disposal method	Incineration	Autoclave ↓ F/b <b>Recycling</b>	Na hypochlorite ↓ Shredding/Pack in puncture proof containers then bury	Na hypochlorite ↓ <b>Recycling</b>

**Sterilization**

00:20:41

Sterilization vs Disinfection :

- Complete killing of all forms of microorganisms
  - Sterilization > Disinfection.
- ↳ Including spore form : Sterilization.  
↳ Excluding spore form : Disinfection.

Types of methods :

	moist Heat	Dry Heat	H <sub>2</sub> O <sub>2</sub> /Plasma	Ethylene oxide (ETO)
Example	Autoclave	Hot air oven	-	-
m/c method	121°C for 15 min under 15 lbs pressure	160°C for 2 hours	-	-
Control	Bacillus Stearothermophilus	Bacillus subtilis/ Bacillus atrophaeus/ Clostridium tetani	Bacillus stearothermophilus	Bacillus subtilis/ Bacillus atrophaeus/ Clostridium tetani

🚮 **materials Sterilized :**

materials	method
<ul style="list-style-type: none"> <li>Surgical instruments except sharps (Eg : Linen, sutures without needle)</li> <li>Plastic &amp; rubber materials (Eg : Syringe, gloves)</li> </ul>	<b>Autoclave</b> > H <sub>2</sub> O <sub>2</sub> > ETO
Glass & sharps (Eg : Flasks, scalpels)	Autoclave > Hot air oven
All culture media (CM)	Autoclave
Exceptions in CM : <ul style="list-style-type: none"> <li><b>Serum</b> CM (Loeffler's serum slope)</li> <li><b>Egg</b> CM (Lowenstein-Jensen medium)</li> </ul>	<b>Inspissation</b> > Tyndallization
Oily & powdery materials (Eg : Liquid paraffin, glove dust powder)	Hot air oven